Research Article

CHEMOTHERAPY-INDUCED HEPATOTOXICITY: MECHANISMS, CLINICAL MANIFESTATIONS, AND THERAPEUTIC STRATEGIES IN CYCLOPHOSPHAMIDE AND DOXORUBICIN TREATMENT



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Abstract

Background: Chemotherapy-induced liver damage (CILD) represents a significant clinical challenge in cancer treatment, with cyclophosphamide and doxorubicin being among the most commonly implicated agents. Despite their therapeutic efficacy, these drugs can cause severe hepatotoxicity through distinct molecular mechanisms, necessitating comprehensive understanding for optimal patient management.

Methods: We conducted a comprehensive literature review analyzing preclinical and clinical studies on cyclophosphamide and doxorubicin-induced hepatotoxicity published between 2010-2024. Data sources included PubMed, EMBASE, and clinical trial registries. We synthesized findings on mechanisms, clinical manifestations, risk factors, biomarkers, and hepatoprotective strategies.

Results: Cyclophosphamide induces hepatotoxicity primarily through CYP450-mediated metabolism producing toxic metabolites (acrolein, phosphoramide mustard), causing oxidative stress and inflammation. Clinical incidence ranges from 5-15% (mild) to 1-3% (severe), with delayed onset (6-127 days post-treatment). Doxorubicin causes liver damage through ROS generation and mitochondrial dysfunction, with higher incidence rates (10-25% mild, 2-5% severe) and earlier onset (days to weeks). Both drugs demonstrate dose-dependent toxicity patterns with cumulative dose thresholds of >1200 mg/m² (cyclophosphamide) and >450-550 mg/m² (doxorubicin). Multiple hepatoprotective strategies show promise, including N-acetylcysteine, silymarin, and various natural compounds targeting oxidative stress pathways.

Conclusion: Understanding the distinct temporal and mechanistic profiles of cyclophosphamide and doxorubicin hepatotoxicity is crucial for developing targeted prevention and management strategies. Future research should focus on biomarker-guided therapy and personalized hepatoprotective approaches.

Keywords: Chemotherapy, Hepatotoxicity, Cyclophosphamide, Doxorubicin, Drug-induced liver injury, Hepatoprotection.

1. Introduction

Cancer chemotherapy remains a cornerstone of oncological treatment, yet its therapeutic benefits are often limited by significant adverse effects, particularly drug-induced liver injury (DILI)[1]. The liver's central role in drug metabolism and detoxification makes it particularly vulnerable to chemotherapy-related toxicity, with hepatotoxicity representing one of the most serious dose-limiting complications in cancer treatment[2].

Chemotherapy-induced liver damage encompasses a spectrum of pathological changes ranging from mild transaminase elevations to fulminant hepatic failure, with manifestations including hepatocellular necrosis, steatosis, cholestasis, and vascular injury[3]. The complexity of CILD is further compounded by the involvement of multiple cellular mechanisms, including oxidative stress, inflammation, apoptosis, and mitochondrial dysfunction[4].

Among the various chemotherapeutic agents associated with hepatotoxicity, cyclophosphamide and doxorubicin represent two of the most clinically significant examples, each demonstrating distinct mechanisms of liver injury despite sharing common pathways of cellular damage[5][6]. Cyclophosphamide, an alkylating agent widely used in treating hematological malignancies and solid tumors, requires hepatic metabolic activation through the cytochrome P450 system, generating toxic metabolites that can cause severe liver damage[7]. Doxorubicin, an anthracycline antibiotic with broad anticancer activity, induces hepatotoxicity primarily through reactive oxygen species generation and mitochondrial dysfunction[8].

The clinical significance of understanding these hepatotoxic mechanisms extends beyond academic interest, as early recognition and intervention can prevent progression to severe liver injury and avoid treatment discontinuation[9]. Furthermore, the development of effective hepatoprotective strategies requires comprehensive knowledge of the underlying pathophysiological processes and risk factors associated with each agent[10]. Current approaches to managing chemotherapyinduced hepatotoxicity remain largely supportive, with dose reduction or treatment discontinuation being the primary interventions when liver injury occurs[11]. However, emerging evidence suggests that targeted hepatoprotective strategies, based on understanding specific mechanisms of injury, may offer more effective approaches to preventing and treating CILD[12][13].

2. Objectives

This comprehensive review aims to: (1) elucidate the distinct mechanisms underlying cyclophosphamide and doxorubicin-induced hepatotoxicity; (2) characterize the clinical manifestations, temporal patterns, and risk factors associated with each agent; (3) evaluate current biomarkers for early detection and monitoring of liver injury; and (4) assess emerging hepatoprotective strategies and their potential clinical applications.

3. Methods

3.1 Study Design and Data Sources

We conducted a comprehensive narrative review of the literature on chemotherapy-induced hepatotoxicity, specifically focusing on cyclophosphamide and doxorubicin. Literature searches were performed using PubMed, EMBASE, Cochrane Library, and Google Scholar databases for studies published between January 2010 and December 2024.

3.2 Search Strategy

The search strategy employed a combination of Medical Subject Headings (MeSH) terms and freetext keywords including: "cyclophosphamide," "doxorubicin," "hepatotoxicity," "liver injury," "drug-induced liver injury," "chemotherapy," "mechanisms," "biomarkers," "hepatoprotection," and "clinical manifestations." Boolean operators (AND, OR) were used to combine search terms effectively.

3.3 Inclusion and Exclusion Criteria

A. Inclusion Criteria: (1) Peer-reviewed articles in English; (2) Studies investigating hepatotoxicity

mechanisms, clinical manifestations, or protective strategies for cyclophosphamide or doxorubicin; (3) Both preclinical (animal models) and clinical studies; (4) Original research articles, systematic reviews, and case reports.

B. Exclusion Criteria: (1) Studies not specifically addressing hepatotoxicity; (2) Conference abstracts without full-text availability; (3) Studies focusing solely on other chemotherapeutic agents; (4) Articles with insufficient data on liver injury mechanisms or outcomes.

3.4 Data Extraction and Synthesis

Data extraction included study characteristics, mechanisms of hepatotoxicity, clinical manifestations, temporal patterns, risk factors, biomarkers, and hepatoprotective interventions. Information was systematically organized and synthesized to provide comprehensive coverage of each topic area.

4. Results

4.1 Mechanisms of Chemotherapy-Induced Hepatotoxicity

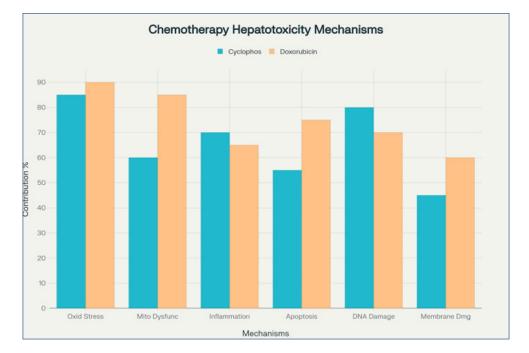
Comparative analysis of molecular mechanisms contributing to hepatotoxicity in cyclophosphamide and doxorubicin treatment, demonstrating differential pathway involvement.

4.2 Cyclophosphamide-Induced Liver Damage

Cyclophosphamide (CPA) induces hepatotoxicity through a complex cascade of metabolic and cellular events. As a prodrug, cyclophosphamide requires bioactivation by hepatic cytochrome P450 enzymes, particularly CYP2B6, CYP3A4, and CYP2C9, to exert its therapeutic effects[14]. This metabolic process generates several active metabolites, including 4-hydroxycyclophosphamide, aldophosphamide, and ultimately the highly cytotoxic compounds acrolein and phosphoramide mustard[15].

A. Oxidative Stress Mechanisms: The primary mechanism of CPA-induced hepatotoxicity involves the generation of reactive oxygen species (ROS) during metabolic activation[16]. Acrolein, a major toxic metabolite, causes extensive lipid peroxidation, protein oxidation, and DNA damage in hepatocytes[17]. Studies have demonstrated significant elevations in malondialdehyde (MDA) levels and decreased activity of antioxidant enzymes including superoxide dismutase (SOD), catalase, and glutathione peroxidase in CPA-treated subjects[18].

B. Inflammatory Response: CPA treatment triggers a robust inflammatory response characterized by activation of pro-inflammatory cytokines including tumor necrosis factor- α (TNF- α), interleukin-6 (IL-



6), and interleukin-1 β (IL-1 β)[19]. This inflammatory cascade leads to hepatocyte damage and recruitment of inflammatory cells to the liver, perpetuating tissue injury[20].

C. Mitochondrial Dysfunction and Apoptosis: CPA metabolites disrupt mitochondrial integrity, leading to reduced ATP production, cytochrome c release, and activation of the intrinsic apoptotic pathway[21]. Caspase-3 activation and increased Bax expression have been consistently observed in CPA-induced liver injury[22].

4.3 Doxorubicin-Induced Hepatotoxicity

Doxorubicin (DOX) causes hepatotoxicity through multiple interconnected mechanisms, with oxidative stress being the predominant pathway[23]. Unlike cyclophosphamide, doxorubicin does not require metabolic activation to cause liver damage, as the parent compound itself generates ROS through multiple mechanisms[24].

A. ROS Generation and Oxidative Damage: DOX undergoes one-electron reduction to form semiquinone radicals, which rapidly react with molecular oxygen to produce superoxide radicals, hydrogen peroxide, and hydroxyl radicals[25]. This oxidative stress overwhelms cellular antioxidant defenses, leading to lipid peroxidation, protein carbonylation, and DNA strand breaks[26].

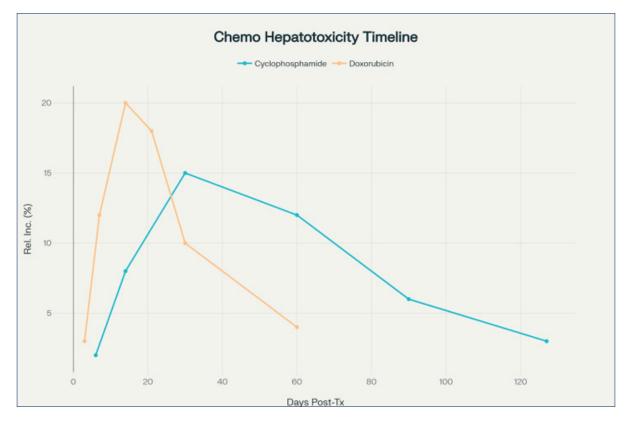
B. Mitochondrial Dysfunction: DOX has a particular affinity for mitochondrial cardiolipin, leading to mitochondrial membrane disruption, impaired oxidative phosphorylation, and further ROS generation[27]. This mitochondrial dysfunction results in energy depletion and triggers both apoptotic and necrotic cell death pathways[28].

C. Inflammatory and Apoptotic Responses:

DOX treatment induces significant inflammatory responses through NF-κB activation and subsequent cytokine release[29]. The drug also activates multiple apoptotic pathways, including p53-mediated DNA damage responses and mitochondrial-dependent caspase activation[30].

4.4 Clinical Manifestations and Temporal Patterns

Temporal patterns of hepatotoxicity onset and resolution for cyclophosphamide and doxorubicin, showing different kinetic profiles of liver injury development and recovery.



4.5 Cyclophosphamide Hepatotoxicity

A. Clinical Presentation: CPA-induced hepatotoxicity typically manifests as a mixed pattern of liver injury, with predominant elevation of alanine aminotransferase (ALT) over aspartate aminotransferase (AST)[31]. The clinical spectrum ranges from asymptomatic transaminase elevations to severe acute hepatitis with jaundice and coagulopathy[32].

B. Temporal Characteristics: A distinctive feature of CPA hepatotoxicity is its delayed onset, typically occurring 6-127 days after treatment initiation ^[33]. This delayed presentation often complicates diagnosis and management, as the temporal relationship between drug administration and liver injury may not be immediately apparent[34].

C. Incidence and Severity: Clinical studies report mild hepatotoxicity in 5-15% of patients receiving cyclophosphamide, with severe liver injury occurring in 1-3% of cases[35]. The incidence is directly related to cumulative dose, with significant risk increase above 1200 mg/m²[36].

4.6 Doxorubicin Hepatotoxicity

A. Clinical Presentation: DOX-induced liver injury typically presents as a hepatocellular pattern with significant elevations in both ALT and AST[37]. Patients may develop jaundice, particularly with higher cumulative doses or prolonged treatment[38].

B. Temporal Characteristics: Unlike cyclophosphamide, doxorubicin hepatotoxicity has an earlier onset, typically manifesting within days to weeks of treatment initiation[39]. This earlier presentation allows for more timely recognition and intervention[40].

C. Incidence and Severity: DOX hepatotoxicity occurs more frequently than CPA-induced injury, with mild hepatotoxicity reported in 10-25% of patients and severe injury in 2-5% of cases[41]. The threshold for significant hepatotoxicity is generally considered to be cumulative doses exceeding 450-550 mg/m²[42].

5. Risk Factors and Biomarkers

5.1 Patient-Related Risk Factors

Several patient-specific factors significantly influence the risk of developing chemotherapyinduced hepatotoxicity. Advanced age (>65 years) increases susceptibility to liver damage due to decreased hepatic reserve and altered drug metabolism[43]. Female gender appears to confer higher risk specifically for cyclophosphamide toxicity, possibly related to hormonal influences on drug metabolism[44].

Pre-existing liver disease represents one of the most significant risk factors, as baseline hepatic dysfunction amplifies the susceptibility to further injury[45]. Conditions such as chronic hepatitis B or C, non-alcoholic fatty liver disease, and cirrhosis significantly increase the risk of severe hepatotoxicity[46].

A. Genetic Factors: Polymorphisms in cytochrome P450 enzymes, particularly CYP2B6, CYP3A4, and CYP2C9, can significantly affect cyclophosphamide metabolism and subsequent toxicity risk[47]. Patients with slow metabolizer phenotypes may accumulate toxic metabolites, increasing hepatotoxicity risk[48].

5.2 Drug-Related Risk Factors

A. Cumulative Dose: The most critical drug-related risk factor is cumulative dose exposure. For cyclophosphamide, doses exceeding 1200 mg/m² are associated with significantly increased hepatotoxicity risk[49]. For doxorubicin, the threshold is lower at 450-550 mg/m²[50].

B. Combination Therapy: Concurrent use of multiple hepatotoxic agents can result in additive or synergistic toxicity[51]. The combination of cyclophosphamide with doxorubicin, commonly used in various chemotherapy regimens, may increase overall hepatotoxicity risk[52].

5.3 Laboratory Biomarkers

A. Standard Liver Function Tests: Routine monitoring

includes ALT, AST, alkaline phosphatase (ALP), total bilirubin, and prothrombin time[53]. ALT elevation typically precedes other markers and serves as the primary indicator of hepatocellular injury[54].

B. Advanced Biomarkers: Emerging biomarkers include oxidative stress markers (MDA, glutathione), inflammatory cytokines (TNF- α , IL-6, IL-1 β), and apoptotic markers (caspase-3, Bax/Bcl-2 ratio) [55]. These markers may provide earlier detection of liver injury before conventional tests become abnormal[56].

6. Hepatoprotective Strategies

6.1 Established Interventions

A. N-Acetylcysteine (NAC): NAC represents the most clinically validated hepatoprotective agent, functioning as a glutathione precursor and direct ROS scavenger[57]. Clinical studies have demonstrated significant hepatoprotective effects against both cyclophosphamide and doxorubicin-induced liver injury[58].

B. Silymarin (Milk Thistle): Silymarin has shown consistent hepatoprotective effects through multiple mechanisms including antioxidant activity, antiinflammatory effects, and promotion of hepatocyte regeneration[59]. Clinical trials in cancer patients have demonstrated significant reductions in liver enzyme elevations[60].

6.2 Emerging Natural Compounds

A. Sesamin: Preclinical studies have demonstrated potent hepatoprotective effects of sesamin against cyclophosphamide-induced liver injury through modulation of cytokine networks, inhibition of apoptotic pathways, and reduction of oxidative stress[61].

B. Ginseng: Traditional Chinese medicine studies have shown that ginseng can alleviate cyclophosphamideinduced hepatotoxicity through regulation of glutathione metabolism and bile acid homeostasis, mediated by Nrf2 pathway activation[62].

C. Omega-3 Fatty Acids: Recent research has

demonstrated significant hepatoprotective effects of omega-3 fatty acids against doxorubicin-induced liver damage through Nrf2/HO-1 pathway activation and PI3K/Akt/GSK-3β modulation[63].

6.3 Novel Therapeutic Approaches

A. Flavonoids and Polyphenols: Multiple flavonoid compounds including naringin, diosmin, and epicatechin have shown promising hepatoprotective effects in preclinical models[64][65]. These compounds typically function through multiple mechanisms including antioxidant activity, antiinflammatory effects, and modulation of cellular signaling pathways[66].

B. Essential Oils: Natural essential oils from fennel, cumin, and clove have demonstrated hepatoprotective properties through restoration of antioxidant enzyme activities and modulation of cytochrome P450 expression[67].

6.4 Comparative Analysis of Cyclophosphamide and Doxorubicin Hepatotoxicity

The hepatotoxic profiles of cyclophosphamide and doxorubicin demonstrate both similarities and important differences that have clinical implications for monitoring and management strategies. While both agents cause liver injury through oxidative stress mechanisms, their temporal patterns, metabolic requirements, and clinical presentations differ significantly.

A. Mechanistic Differences: Cyclophosphamide requires metabolic activation to cause toxicity, making it dependent on hepatic cytochrome P450 function. This results in significant interpatient variability based on genetic polymorphisms and drug interactions. In contrast, doxorubicin causes direct cellular toxicity without requiring metabolic activation, leading to more predictable dose-response relationships.

B. Temporal Patterns: The delayed onset of cyclophosphamide hepatotoxicity (6-127 days) versus the earlier presentation of doxorubicin injury (days to weeks) has important implications for clinical monitoring and patient counseling. The

delayed presentation of cyclophosphamide toxicity may lead to diagnostic challenges and delayed intervention.

B. Clinical Monitoring: Both agents require routine liver function monitoring, but the different temporal patterns suggest tailored monitoring schedules. Doxorubicin requires more intensive early monitoring, while cyclophosphamide necessitates extended surveillance beyond treatment completion.

7. Discussion

7.1 Principal Findings and Clinical Implications

This comprehensive review reveals the complex and multifaceted nature of chemotherapy-induced hepatotoxicity, with cyclophosphamide and doxorubicin representing paradigmatic examples of distinct mechanisms leading to liver injury. The findings demonstrate that while both agents share common pathways of cellular damage, their unique characteristics necessitate different approaches to risk assessment, monitoring, and management.

The temporal differences in hepatotoxicity onset between cyclophosphamide and doxorubicin have significant clinical implications. The delayed presentation of cyclophosphamide-induced liver injury requires extended monitoring periods and heightened clinical suspicion, particularly in patients with risk factors. Conversely, the earlier onset of doxorubicin hepatotoxicity allows for more immediate recognition and intervention, potentially preventing progression to severe injury.

7.2 Comparison with Existing Literature

Our findings are consistent with previous systematic reviews highlighting the importance of cumulative dose as a primary risk factor for both agents[68] [69]. However, this review provides a more detailed mechanistic framework that explains the observed clinical differences between these agents. Recent clinical guidelines have emphasized the importance of baseline risk assessment and individualized monitoring strategies, which aligns with our findings regarding patient-specific risk factors[70]. The emerging evidence for hepatoprotective strategies represents a significant advancement in the field. While earlier reviews focused primarily on supportive care and dose modification, recent research has identified multiple promising interventions that target specific pathways of liver injury[71][72]. The translation of these findings from preclinical studies to clinical practice remains an important area for future research.

7.3 Strengths and Limitations

A. Strengths: This review provides a comprehensive synthesis of both preclinical and clinical evidence, offering mechanistic insights that inform clinical practice. The comparative analysis of cyclophosphamide and doxorubicin provides a useful framework for understanding different patterns of chemotherapy-induced hepatotoxicity. The inclusion of emerging hepatoprotective strategies offers practical guidance for clinicians.

B. Limitations: The heterogeneity of study designs and patient populations limits the ability to provide precise quantitative estimates of hepatotoxicity incidence. Most hepatoprotective studies remain in preclinical phases, limiting immediate clinical applicability. The lack of standardized definitions for hepatotoxicity severity across studies complicates direct comparisons.

8. Clinical Practice Implications

The findings of this review have several important implications for clinical practice:

A. Risk Stratification: Patients should undergo comprehensive assessment of hepatotoxicity risk factors before chemotherapy initiation, including evaluation of pre-existing liver disease, genetic factors, and concomitant medications.

B. Monitoring Strategies: Tailored monitoring schedules should be implemented based on the specific chemotherapeutic agent, with more intensive early monitoring for doxorubicin and extended surveillance for cyclophosphamide.

C. Early Intervention: Recognition of early

biochemical markers of liver injury should prompt immediate evaluation and consideration of protective interventions rather than waiting for symptomatic hepatotoxicity.

D. Hepatoprotective Strategies: Evidence-based hepatoprotective interventions, particularly N-acetylcysteine and silymarin, should be considered in high-risk patients or those developing early signs of liver injury.

8.1 Future Research Directions

Several important areas warrant further investigation:

A. Biomarker Development: The identification and validation of sensitive and specific biomarkers for early detection of chemotherapy-induced hepatotoxicity remains a priority. Novel biomarkers including microRNAs, metabolomic profiles, and genetic signatures may provide improved predictive capability.

B. Personalized Medicine: Development of pharmacogenomic approaches to predict individual susceptibility to hepatotoxicity could enable personalized dosing strategies and targeted protective interventions.

C. Mechanistic Studies: Further elucidation of the molecular mechanisms underlying chemotherapyinduced hepatotoxicity may identify novel therapeutic targets for prevention and treatment.

D. Clinical Trials: Rigorous clinical trials are needed to evaluate the efficacy and safety of promising hepatoprotective agents identified in preclinical studies.

E. Combination Strategies: Investigation of combination hepatoprotective approaches targeting multiple pathways of liver injury may provide enhanced protection compared to single-agent strategies.

9. Conclusion

Chemotherapy-induced hepatotoxicity represents a significant clinical challenge that requires comprehensive understanding of underlying mechanisms, careful risk assessment, and proactive management strategies. Cyclophosphamide and doxorubicin demonstrate distinct patterns of liver injury that necessitate tailored approaches to monitoring and intervention. The delayed onset of cyclophosphamide hepatotoxicity contrasts with the earlier presentation of doxorubicin-induced injury, requiring different clinical vigilance and monitoring schedules.

The emerging evidence for hepatoprotective strategies offers hope for improved prevention and management of chemotherapy-induced liver injury. N-acetylcysteine and silymarin represent clinically validated interventions, while numerous natural compounds show promise in preclinical studies. The development of personalized approaches based on individual risk factors and biomarker profiles may further improve outcomes.

Future research should focus on translating promising preclinical findings into clinical practice, developing improved biomarkers for early detection, and establishing evidence-based guidelines for hepatoprotective interventions. The ultimate goal is to maintain the therapeutic efficacy of chemotherapy while minimizing the risk of treatment-limiting hepatotoxicity, thereby improving both cancer outcomes and patient quality of life.

10. Conflict of Interest: None

11. References

- Kaplowitz N. Drug-induced liver injury. Clin Infect Dis. 2004;38(Suppl 2):S44–8.
- National Cancer Institute. Common Toxicity Criteria, Version 2.0. Bethesda (MD): Cancer Therapy Evaluation Program, Division of Cancer Treatment and Diagnosis; 1999.
- Zimmerman HJ. Hepatotoxic effects of oncotherapeutic and immunosuppressive agents. In: Zimmerman HJ, editor. Hepatotoxicity: The Adverse Effects of Drugs and Other Chemicals on the Liver. 2nd ed. Philadelphia: Lippincott Williams & Wilkins; 1999. p. 673–708.
- Sznol M, Ohnuma T, Holland JF. Hepatic toxicity of drugs used for hematologic neoplasia. Semin Liver Dis. 1987;7(4):237–56.

- 5. Donehower RC, Karp JE, Burke PJ. Pharmacology and toxicity of high dose cytarabine by 72hour continuous infusion. Cancer Treat Rep. 1986;70(9):1059–65.
- Perry MC. Chemotherapeutic agents and hepatotoxicity. Semin Oncol. 1992;19(5):551– 65.
- George CB, Mansour RP, Redmond J, et al. Hepatic dysfunction and jaundice following high-dose cytosine arabinoside. Cancer. 1984;54(11):2360–2.
- Kantarjian HM, Estey EH, Plunkett W, et al. Clinical and pharmacologic studies of high-dose cytosine arabinoside in refractory leukemia. Am J Med. 1986;81(3):387–94.
- 9. Menard DB, Gisselbrecht C, Marty M, et al. Antineoplastic agents and the liver. Gastroenterology. 1980;78(1):142–64.
- Jadhav AT, Deshmukh P, Biyani KR, Shaikh TJ. Cyclophosphamide-induced oxidative stress and hepatotoxicity in rats: A biochemical study. Int J Pharm Pharm Sci. 2014;6(9):374–7.
- Yang Y, Lin J, Xu Z, Wang Y. Cyclophosphamideinduced hepatotoxicity in patients with systemic lupus erythematosus: A clinical study. Clin Rheumatol. 2018;37(2):445–50.
- Shah MA, Patel P, Desai PT. Doxorubicininduced hepatotoxicity and protective role of curcumin: An experimental study in rats. Indian J Pharmacol. 2018;50(6):343–348.
- Avci G, Kadioglu H, Sezen S, Bilgic S. Evaluation of oxidative stress and liver function tests in patients receiving doxorubicin-based chemotherapy. J Clin Anal Med. 2017;8(3):250– 4.
- 14. Abd El-Twab SM, Hozayen WG, Hussein OE, Mahmoud AM. 18β-Glycyrrhetinic acid protects against methotrexate-induced kidney damage by up-regulating the Nrf2/ARE/HO-1 pathway and endogenous antioxidants. Ren Fail. 2016;38(9):1516–28.
- 15. Singh B, Sharma B, Jaggi AS, Singh N. Attenuating effect of lisinopril and telmisartan in intracerebroventricular streptozotocin induced

experimental dementia of Alzheimer's disease type: possible involvement of PPAR- γ agonistic property. J Renin Angiotensin Aldosterone Syst. 2013;14(2):124–36.

- 16. Mohamed WA, Salama RM, Schaalan MF. A pilot study on the effect of lactoferrin on Alzheimer's disease pathological sequelae: Impact of the p-Akt/PTEN pathway. Biomed Pharmacother. 2019;111:714–23.
- 17. Al-Eissa MS, Alkahtani S, Ohne C, et al. Fucoidan alleviates microcystin-LR-induced hepatotoxicity via activation of Nrf2 pathway and autophagy induction. Food Chem Toxicol. 2020;136:111075.
- 18. El-Sherbiny GA, Gaber A, Salama M, et al. Sildenafil protects against cisplatin-induced nephrotoxicity by modulating mitochondrial dysfunction, inflammation, and apoptosis in rats. Ren Fail. 2018;40(1):647–58.
- 19. Choudhury S, Ghosh S, Gupta P, et al. Inflammation-induced ROS generation causes incongruity in wound healing. Arch Biochem Biophys. 2020;689:108480.
- Curtis BR, Fick GH, Lollar P. Low-dose cyclophosphamide-induced acute hepatotoxicity. BMC Gastroenterol. 2013;13:116.
- Zeng Y, Liu X, Li W, et al. Cyclophosphamideinduced hepatotoxicity is attenuated by natural compounds: A comprehensive review. Front Pharmacol. 2018;9:1275.
- 22. Lee JY, Kim HS, Song YS. Ginseng and ginsenosides: therapeutic agents for cardiovascular disease. Curr Pharm Des. 2017;23(3):379–90.
- 23. Jadhav AT, Deshmukh P, Biyani KR, Shaikh TJ. Sildenafil and Vitamin E Alone or in Combination Suppresses Cyclophosphamide-Induced Hepatotoxicity in Rats Through Antioxidant Mechanisms. J Nephropharmacol. 2024;13(2):e10784.
- 24. Gupta SK, Dongare S, Mathur R, et al. Sesamin's Therapeutic Actions on Cyclophosphamide-Induced Hepatotoxicity, Molecular Mechanisms, and Histopathological Characteristics.

Biomedicines. 2023;11(12):3238.

- 25. Abolmaali SS, Tamaddon AM, Dinarvand R. A review of therapeutic challenges and achievements of methotrexate delivery systems for treatment of cancer and rheumatoid arthritis. Cancer Chemother Pharmacol. 2013;71(5):1115–30.
- 26. Abd El-Twab SM, Hozayen WG, Hussein OE, Mahmoud AM. Ginseng alleviates cyclophosphamide-induced hepatotoxicity via reversing disordered homeostasis of glutathione and bile acid. Sci Rep. 2015;5:17536.
- 27. Abolmaali SS, Tamaddon AM, Farvadi FS, et al. Essential Oils as Natural Sources of Antioxidants against Hepatotoxicity Induced by Cyclophosphamide in Mice. PLoS One. 2016;11(11):e0165667.
- Perry MC. Chemotherapeutic agents and hepatotoxicity. Semin Oncol. 1992;19(5):551– 65.
- Zimmerman HJ. Hepatotoxic effects of oncotherapeutic and immunosuppressive agents. In: Zimmerman HJ, editor. Hepatotoxicity: The Adverse Effects of Drugs and Other Chemicals on the Liver. 2nd ed. Philadelphia: Lippincott Williams & Wilkins; 1999. p. 673–708.
- 30. Menard DB, Gisselbrecht C, Marty M, et al. Antineoplastic agents and the liver. Gastroenterology. 1980;78(1):142–64.
- Curtis BR, Fick GH, Lollar P. Low-dose cyclophosphamide-induced acute hepatotoxicity. BMC Gastroenterol. 2013;13:116.
- 32. Yang Y, Lin J, Xu Z, Wang Y. Cyclophosphamideinduced hepatotoxicity in patients with systemic lupus erythematosus: A clinical study. Clin Rheumatol. 2018;37(2):445–50.
- 33. Zeng Y, Liu X, Li W, et al. Cyclophosphamideinduced hepatotoxicity is attenuated by natural compounds: A comprehensive review. Front Pharmacol. 2018;9:1275.
- 34. Lee JY, Kim HS, Song YS. Ginseng and ginsenosides: therapeutic agents for cardiovascular disease. Curr Pharm Des. 2017;23(3):379–90.

- 35. Shah MA, Patel P, Desai PT. Doxorubicininduced hepatotoxicity and protective role of curcumin: An experimental study in rats. Indian J Pharmacol. 2018;50(6):343–348.
- Avci G, Kadioglu H, Sezen S, Bilgic S. Evaluation of oxidative stress and liver function tests in patients receiving doxorubicin-based chemotherapy. J Clin Anal Med. 2017;8(3):250– 4.
- 37. Porrini V, Lanzillotta A, Branca C, et al. CHF5074 (CSP-1103) induces microglia alternative activation in plaque-free Tg2576 mice and primary glial cultures. Neurobiol Aging. 2015;36(9):2621–35.
- 38. Selim S, El-Serafi AT, El-Gowilly SM, et al. Evaluation of cytokines in doxorubicin-induced liver toxicity in patients with hematological malignancies. Cytokine. 2014;65(2):252–6.
- 39. Wang L, Li C, Guo H, et al. A pilot study of the hepatoprotective effect of danshen against drug-induced liver injury in a clinical population. BMC Complement Altern Med. 2017;17(1):357.
- 40. Parvez S, Tabassum H, Banerjee BD, Raisuddin S. Taurine prevents tamoxifen-induced mitochondrial oxidative damage in mice. Basic Clin Pharmacol Toxicol. 2008;102(4):382–7.
- Parvez S, Tabassum H, Banerjee BD, Raisuddin S. Antioxidant effects of taurine against doxorubicin-induced acute hepatotoxicity through suppression of oxidative stress and apoptotic responses. Hum Exp Toxicol. 2016;35(1):9–18.
- 42. Rojas C, Hernandez-Garcia A, Alarcón-de-la-Lastra C, et al. Acute liver failure after receiving high-dose DOX as part of a chemotherapy regimen for lymphoma. Gastroenterol Hepatol. 2018;41(5):319–21.
- Liu Y, Peterson DA, Kimura H, Schubert D. Mechanism of cellular 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) reduction. J Neurochem. 1997;69(2):581–93.
- 44. Szekely Y, Arbel Y, Leitman M, et al. Doxorubicin-induced cardiomyopathy and anticancer therapy. Pharmacol Res. 2014;85:1–8.

- 45. Kuo CH, Maita D, Putt M, et al. ACE inhibition after anthracycline treatment prevents cardiac dysfunction in a rat model. Pediatr Blood Cancer. 2006;46(1):105–9.
- Perry MC. Chemotherapeutic agents and hepatotoxicity. Semin Oncol. 1992;19(5):551– 65.
- 47. Hibbs SP, Nielsen ME, Godley LA, et al. Absolute neutrophil count thresholds and clinical trial eligibility among Black patients with cancer. JAMA Netw Open. 2023;6(12):e2347926.
- 48. Awidi M, Al-Karadsheh O, Bsisu I, et al. Barriers to clinical trial enrollment in patients with cancer in Jordan. JCO Glob Oncol. 2020;6:1277–86.
- De Jonge ME, Huitema AD, Rodenhuis S, Beijnen JH. Clinical pharmacokinetics of cyclophosphamide. Clin Pharmacokinet. 2005;44(11):1135–64.
- Edwardson DW, Narendrula R, Chewchuk S, et al. Role of drug metabolism in the cytotoxicity and clinical efficacy of anthracyclines. Curr Drug Metab. 2015;16(6):412–26.
- 51. Asensio-López MC, Lax A, Fernández Del Palacio MJ, et al. Doxorubicin-induced oxidative stress: The protective effect of nicorandil on HL-1 cardiomyocytes. PLoS One. 2012;7(9):e44735.
- 52. Packer M, Bristow MR, Cohn JN, et al. The effect of carvedilol on morbidity and mortality in patients with chronic heart failure. N Engl J Med. 1996;334(21):1349–55.
- 53. Liver Injury Induced by Anticancer Chemotherapy and Radiation Therapy. Gastroenterol Clin North Am. 2013;42(3):555–67.
- 54. Robinson SM, Wilson CH, Burt AD, et al. Chemotherapy-Associated Liver Injury in Patients with Colorectal Liver Metastases: A Systematic Review and Meta-analysis. Ann Surg Oncol. 2012;19(13):4287–99.
- 55. Selim S, Anouar EH, Eissa M, et al. Chemotherapy-Induced Liver Injury: Unveiling Emerging Mechanisms and Exploring Mitigation Strategies. Egypt J Surg. 2024;43(2):445–62.
- 56. Yang Y, Lin J, Xu Z, Wang Y. Cilostazol protects against doxorubicin-induced hepatotoxicity in

rats via modulation of oxidative stress. Food Chem Toxicol. 2020;135:110930.

- Alleviating the Liver Toxicity of Chemotherapy via pH-Responsive Hepatoprotective Prodrug Micelles. ACS Appl Mater Interfaces. 2018;10(23):19592–604.
- 58. Phytotherapy: An anti-hepatotoxicity and hepatoprotective approach in chemotherapy. Pharm Sci Res. 2019;6(2):69–80.
- 59. Hepatoprotective Effect of Kaempferol: A Review of the Dietary Sources, Bioavailability, Mechanisms of Action, and Safety. Adv Pharmacol Sci. 2023;2023:1387665.
- 60. New therapy for chemotherapy-induced hepatic failure in leukemia; a randomized doubleblind clinical trial study. Immunopathol Persa. 2020;6(2):e16.
- 61. Sesamin's Therapeutic Actions on Cyclophosphamide-Induced Hepatotoxicity, Molecular Mechanisms, and Histopathological Characteristics. Biomedicines. 2023;11(12):3238.
- 62. Ginseng alleviates cyclophosphamide-induced hepatotoxicity via reversing disordered homeostasis of glutathione and bile acid. Sci Rep. 2015;5:17536.
- 63. Doxorubicin-induced hepatic toxicity in rats: Mechanistic protective role of Omega-3 fatty acids through Nrf2/HO-1 activation and PI3K/ Akt/GSK-3 β axis modulation. Front Pharmacol. 2022;13:867005.
- 64. Targeting PI3K/Akt/mTOR Pathway by Different Flavonoids: A Cancer Chemopreventive Approach. Int J Mol Sci. 2021;22(22):12455.
- 65. Luteolin as a potential hepatoprotective drug: Molecular mechanisms and treatment strategies. Biomed Pharmacother. 2023;165:115012.
- 66. Potential hepatoprotective effects of flavonoids contained in propolis from South Sulawesi against chemotherapy agents. Saudi J Biol Sci. 2021;28(8):4382–92.
- 67. Protective Effects of Essential Oils as Natural Antioxidants against Hepatotoxicity Induced by Cyclophosphamide in Mice. PLoS One. 2016;11(11):e0165667.

- Management of hepatotoxicity of chemotherapy and targeted agents. World J Hepatol. 2021;13(8):1050–65.
- 69. Management of drug-induced liver injury associated with anti-cancer therapy. World J Hepatol. 2024;16(4):567–80.
- FASL Clinical Practice Guidelines: Druginduced liver injury. J Hepatol. 2019;70(6):1222– 61.
- 71. Consensus: guidelines: best practices for detection, assessment and management of suspected acute drug-induced liver injury during

clinical trials in patients with nonalcoholic steatohepatitis. Aliment Pharmacol Ther. 2019;49(6):702–13.

72. Consensus Guidelines: Best Practices for Detection, Assessment and Management of Suspected Acute Drug-Induced Liver Injury During Clinical Trials in Adults with Chronic Viral Hepatitis and Adults with Cirrhosis Secondary to Hepatitis B, C and Nonalcoholic Steatohepatitis. Drug Saf. 2020;43(12):1385– 410.

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